



PHYTOCHEMICAL ANALYSIS OF SIDA OVATA SEED OIL: A NEW SOURCE OF CYCLOPROPENOID FATTY ACID

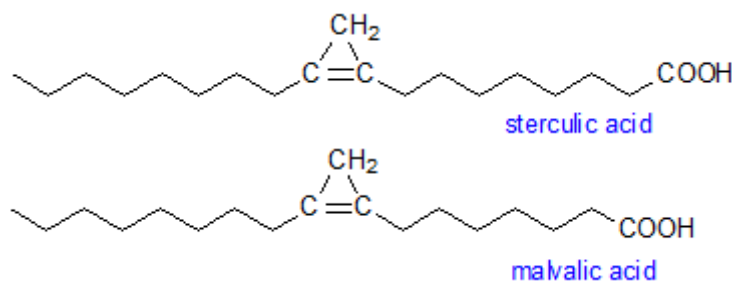
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ABSTRACT

The seeds of *Sidaovata* were collected from the arid zone of Rajasthan and analysed for the characterization of the fatty acids. This paper accounted the presence of Cyclopropenoid fatty acids (Malvalic and Sterculic). The occurrence of CPFA was confirmed by HBr-titration, Halphen test, TLC, NMR and IR spectroscopic methods.

Keywords: *Sidaovata* (Malvaceae), seed oil characterization, Cyclopropenoid fatty acids.



Structure of Malvalic and Sterculic acids (Cyclopropenoid fatty acids)

INTRODUCTION

The plant of Sidaovata (Malvaceae) is widely spread in the arid parts of tropical land in India. The roots of Sida species are considered excellent adaptogenic and immunomodulator, general nutritive tonic and prolonged life, useful in tuberculosis, heart diseases, cough and respiratory diseases¹. The plant species belonging to Malvaceae and Sterculiaceae are growing abundantly throughout the Arid zone of Rajasthan and all over India, the fruit of some species are demulcents, mildly astringent and commonly used in griping and flatulence².

The seed oil of Sidaovata responded positive Halphen test³ and was selected for the estimation and characterization of CPFA. The naturally occurring CPFA are sterculic (n=7) and malvalic (n=6), found mainly in the seed lipids of four plant families of order malvales which includes (Sterculiaceae, Malvaceae, Bombaceae and Tiliaceae). CPFA occur in plant lipids mainly as glycerides. CPFA have been the subject of many investigations due to their Carcinogenic, Co-Carcinogenic activities⁴⁻⁸. The presence of CPFA in food is dangerous not only for human health but also for animals⁹⁻¹⁴. Quantitative estimation of total cyclopropenoid fatty acids content can be achieved most conveniently by HBr titration to a level of .01%¹⁵.

EXPERIMENTAL

The Soxhlet-extraction method was adopted to extract oil from ground seeds, using petroleum ether (40-60°C). The moisture contents, Iodine value, Saponification value of the oil were estimated by standard AOCS methods¹⁶. The protein content of the oil was estimated by Kjeldahl method. Thin layer chromatographic techniques were used to resolve different fatty acid components of the oil along with silver ion TLC¹⁷.

The spots were made visible on analytical plates by charring with 20% aqueous solution of perchloric acid. The transesterification of the oil was carried out using sodium methoxide. Preparative TLC were used for the separation of large quantity of cyclopropenoid fraction. AmilNucon gas chromatograph model no. 5700 equipped with a flame ionization detector was used for the GLC study of oil samples. The U.V spectra of the oil and methyl esters have been conducted with a Shimadzu UV-1601 spectrophotometer. The NMR spectra were recorded on Bruker X300 spectrophotometer and oil samples were prepared in CDCl₃ containing tetramethylsilane as an internal standard. The IR spectra of oil were obtained from Jasco-made FT-IR spectrophotometer.

RESULTS AND DISCUSSION

The seed oil Sidaovata was evaluated for its physiochemical properties (Table-I). The quantitation of total CPFA material was carried out by H-Br titration which revealed the presence of 16% by weight of CPFA¹⁵. Thin layer liquid chromatography revealed two spots. The methyl ester obtained from the oil sample exhibited the IR band at 1010cm⁻¹ and 1852cm⁻¹, supporting the presence of cyclopropene ring. The characterization of unusual acids was achieved by separation using preparative TLC.

Characterization of fraction-I:

The combined GLC-TLC techniques were used for the analyses of component acids in the CPFA fraction. This fraction have R_F value 0.75 on TLC. The quantitative results of direct, reversed phase and silver ion TLC of this fraction were compared with the esters of Sterculiafoetida used as a reference standard. The spots corresponding to saturated, monoene, diene and triene in this fraction of oil were resolved by AgNO₃-TLC[17] parallel to those obtained from Linseed esters. The negative Halphen test indicated the absence of CPFA in this fraction. The IR spectrum did not show any peak corresponding to CPFA, however a band at 1710cm⁻¹ observed corresponding to ester-carbonyl group.

Characterization of fraction-II:

This fraction showed R_F 0.55. The occurrence of CPFA in this fraction was confirmed by the positive Halphen test[3]. The resulting methyl esters obtained from the base catalyzed trans-esterification showed IR band at 1010cm⁻¹ and 1852cm⁻¹. The quantitative estimation of CPFA by HBr-titration showed the presence of 16% by wt. of CPFA. The absence of conjugation or trans-unsaturation in this fraction of oil was confirmed by U.V spectrum. The NMR spectra of this fraction showed typical cyclopropenoid group signal at δ 0.71 (singlet) other proton signals were also observed at δ 3.6 (3H, -COCH₃), δ 2.2 (2H, α to carbonyl), δ 1.2 (chain-CH₂-) and δ 0.88 (3H, term-CH₃). The GLC analysis of the AgNO₃-methanol treated methyl esters confirmed the presence of Malvalic and Sterculic acids by comparison of relative retention time of similar derivatives from Sterculiafoetida esters. The GLC data given in table-II of the CPFA were found in close agreement with those obtained by H-Br titration [15] methods. The Fatty acids composition of the seed oil is shown in Table-II. The Malvalic and Sterculic acids were found at levels of 14.20% and 6.30% respectively as silver nitrate adduct and the products obtained have been depicted in the scheme. The presence of CPFA has been reported for the first time in the seed oil of Sidaovata.

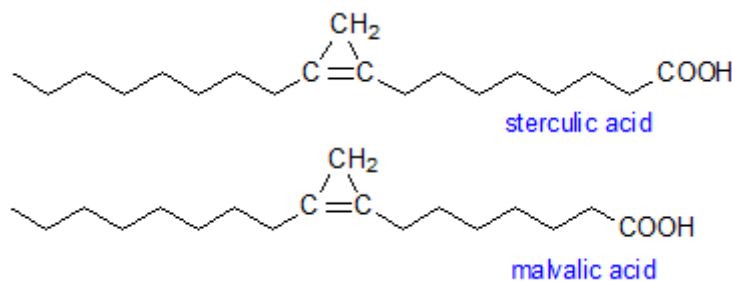
Name and Family	16:0	18:0	18:1	18:2	18:3	20:0	Others
Sida ovate (Malvaceae)	35.2	12.3	15.9	29.6	6.9	-	Traces

Table 2: Fatty Acid Composition Determined by GLC of Fraction-I

Fatty Acids	Sterculiafoetida	Fraction-II
Myristic	-----	2.4
Palmitic	24.8	18.0
Palmitoleic	1.0	-----
Stearic	3.4	2.4
Oleic	9.4	14.6
Linoleic	1.8	38.5
Linolenic	1.2	3.5
Malvalic	7.1	14.2
Sterculic	51.2	6.3

Table 3: GLC Analysis of AgNO₃ –Methanol treated Methyl Esters:

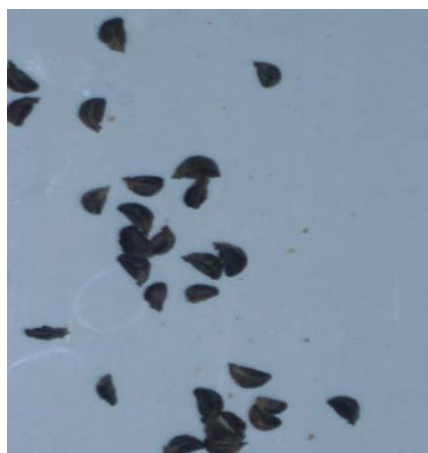
Fraction-II (uncorrected wt. %)



Structure of Malvalic and Sterculic acids



Sidaovata plant



Sidaovata seeds

REFERENCES

1. Knowledge repository- Central institute of Medicinal and Aromatic plants.
2. Kirthikar, K.R and Basu, B.D. An ICS.Indian medicinal plants, VI,II, Lalit Mohan Basu, Allahabad, India,1981;370-372.
3. Halphen, G. J.Pharm. 1897;6 (6th series), 390.
4. Hendrick, J.D; Sinnhuber, R.O; Loveland, P.M; Pawlowski, N.E; Nixon, J.E. Hepatocarcinogenicity of glandless cotton seede and cotton seedoil to rainbow trout (*Salmogairdnerii*). Science 1980; 208:309-311.
5. Sinnhuber, R.O; Lee, D.J; Wales, J.H; Landens, M.K; Keyl, A.C. Hepatic carcinogenesis of alfatoxin M, in rainbow trout (*Salmogairdnerii*) and its enhancement by cyclopropene Fatty acids. Cancer Res, 1974; 53: 1285-1291.
6. Sinnhuber, R.O; Lee, D.J; Wales, J.H; Ayres, J.L. Dietary factors and hepatoma in rainbow trout (*Salmogairdnerii*).2. Carcinogenesis by cyclopropenoidfaty acids and effects of gossypol and altered lipids on alfatoxin-induced liver cancer. J Natt cancer Inst.1968; 41: 1293-1299.
7. Lee, D.J; Wales, J.H; Landens; Sinnhuber, R.O. Hepatoma and renal tubule adenoma in rats fed alfatoxin and cyclopropenoid fatty acids. J Natl cancer Inst.1969; 43; 1037-1044.
8. Lee, D.J; Wales, J.H; Sinnhuber; R.O. Promotion of alfatoxin-induced hepatoma growth in trout by methyl malvalate and stercolate cancer. Res 1971; 31: 960-969.
9. Tumbelaka, L.I; Slayden, O.V; Stormshak; F. Action of cyclopropenoidfattyacid on the corpus luteum of pregnant and non-pregnant eves. BiolReprod. 1994; 50:25. 3-257.
10. Andrianaivo-Rafehivola, A.A; Siess, M.H; Gaydou, E.M. Modifications of hepatic drug metabolizing enzyme activities in rats feb baobab seed oil cyclopropenoid fatty acids. Food. Chem. Toxicol. 1995;33:337-382.

11. Matlock, J.P; Nixon, J.E and Pawlowski, N.E. Altered lipid metabolism and impaired clearance of plasma cholesterol in mice fed cyclopropenoid fatty acids. *ToxicolApplPharmacol* 1985;80: 457-466.
12. Artman, N.R. The chemical and biological properties of heated and oxidized fats. *Adv. Lipid.Res.*1969; 7: 245-330.
13. Nolen, G.A; Alexander, J.C and Artman, N.R. Long term rat feeding study with used frying fats. *J.Nutr* 1967; 93:337-349.
14. Shone, G.G. Chemistry Department, North Staffordshire college of Technology, Stoke-on-Trent.
15. Harris, J.A; Magne, F.C and Skau, E.L. *J. AMER.Oil Chem. Soc.*,1964;4:309.
16. *Official and Tentative Methods of the American Oil Chemists Society*, 1978; 3rdEd..
17. Schneider, E.L; Loke, S.P and Hopkins, D.T. *J. Amer.OilChem.Soc.*, 1968;45:585.